



## **Bacteriological and Physicochemical Assessment of Water from Student Hostels of Osun State University, Main Campus, Osogbo, Southwest Nigeria**

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**Abstract:** *This study was conducted to investigate the potability of 15 samples of water, three each from boreholes (BH1-BH3) and hand-dug wells (HD1-HD3), and nine brands of sachet water (SW1-SW9) that were regularly patronized by the students of Osun State University, main campus, residing in private hostels in Osogbo metropolis. The objective of the study was to determine the quality of such water samples. Borehole and well water samples from selected areas and samples of sachet water regularly vended by different manufacturers and vendors were collected, and subjected to physical, chemical and bacteriological analysis. For physical and chemical analysis Wagtech's photometer plus chemical reagents were used. Total heterotrophic bacteria, total coliforms, and fecal coliforms in the water samples were obtained using, respectively, the pour plate method, membrane filtration and growth on MacConkey*

agar as well as Eosin Methylene Blue agar. The results showed that all of the samples of sachet water exhibited values of physical / organoleptic parameters, inorganic constituents, and mean coliform and *E. coli* counts per 100 ml below the WHO/SO maximum permissible levels; and were therefore, considered safe for drinking. There were slightly elevated levels of iron in water samples from the borehole category, BH2 and BH3 with no known health impacts. This is because iron is an essential element in human nutrition. Taste is not usually noticeable at iron concentrations below 0.3 mg/l. Although iron concentrations of 1–3 mg/l can be acceptable for people drinking well-water, no health-based guideline value for iron has been proposed. However, there were slightly elevated nitrate levels in samples from hand-dug wells, HD1 and HD3, suggesting that these water sources were not safe for consumption by infants under three months old. In addition, evidence of faecal coliform in water samples from hand-dug wells HD1 and HD2 suggest that they were not safe for human drinking. It is recommended that water from hand dug wells should be boiled before consumption to ensure public health and safety.

## **Introduction**

Water is essential to sustain life, and a satisfactory supply must be made available to consumers. Potable water is water pure enough to be consumed or used with low risk of immediate or long term harm. Like other developing countries, the issue of access to potable water is very important in Nigeria, where 48% (about 67 million Nigerians) depend on surface water for domestic use, 57% (79 million) use hand dug wells, 20% (27.8 million) harvest rain, 14% (19.5 million) have access to pipe borne water, and 14% have access to borehole water sources (FGN, 2007). The quality and quantity of pipe borne water for drinking is deteriorating in the country due to inadequacy of treatment plants, direct discharge of untreated sewage into rivers and streams, and inefficient management of piped water distribution system (UNEP, 2001; Aderibigbe *et al.*, 2008; Gbadegesin and Olorunfemi,

2007). Although, the importance and use of groundwater has increased significantly during the last decades in urban and rural areas of the country (Adekunle *et al.*, 2007), contamination from natural and human activities is affecting this vital resource (Adetunde *et al.*, 2011; Ishaku *et al.*, 2011). A recent local intervention, drinking water sold in polythene sachets, though readily available and affordable, has generated public health concerns about purity due to the questionable integrity of the production environments and contamination from improper vendor handling (Adekunle *et al.*, 2004; Dada, 2008).

Water which is not safe to drink can carry pathogenic microbes and heavy metals. Globally, about 80% of all diseases and death in developing countries are water-related as a result of polluted water (Ayeni *et al.*, 2011, Aderibigbe *et al.*, 2008). The numbers of water-borne disease outbreaks that have been reported in

Nigeria demonstrate that transmission of pathogens by drinking water remains a significant cause of illness (Nwidi *et al.*, 2008). Although, the symptoms of gastrointestinal disorders (nausea, diarrhea, vomiting and abdominal pain) are usually mild and generally last a few days to a week and only a small percentage of those affected will visit a health facility, there is untold impact on human productivity.

Osun State University is a non-residential institution of higher learning with six campuses distributed across Osun State, Southwest of Nigeria. The main campus of the University is situated in Oke Baale area of Osogbo, the state capital. Students live in private hostels near the main campus and other communities in the metropolis. The water supplies from the Ministry of Water Resources and Rural Development, Osun State Water Corporation (OSWC), and the Rural Water Environmental Sanitation Agency (RUWESA) are inadequate for the growing population and at best epileptic in most Osogbo communities (OSWC, 2008). As a consequence of pipe borne water shortage, the Osun State University students depend on water from boreholes, hand-dug wells and water vended in sachets. From a public health perspective, there is a need to ascertain the quality of water accessible to the university student hostels to avoid or reduce incidence

of contaminated water-related health hazards. Based on this, the study was undertaken to assess the physical, chemical, and bacteriological properties of the water sources used by the students of Osun State University main campus. The study is relevant in assuring whether the quality of the water sources meet regulatory standards and is significant because potable water is essential to life.

## **Materials and Methods**

### **Study Area**

The study was conducted in Osogbo, a commercial and industrial city (located 7°46' North 4°34' East with an area of 47km sq), and the capital of Osun State, Nigeria. Specifically, private hostels at Oke Baale, Abe Obi, Kasmu, and Ogo Oluwa communities of Osogbo occupied by the students of Osun State University, main campus, were used in the present study.

### **Water Collection**

A total of 15 samples of water, three each from hand dug-wells and boreholes, and nine brands of sachet water vended by different manufacturers and vendors were collected between December 2011 and February 2012, during the dry season of the year when the problem of water access and supply was very acute. Three replicates were collected per source. Well water sample was collected directly from each well by means of a sterile plastic container fitted with a weight

at the base; taking care to avoid contamination by surface scum. The outlet of each borehole was disinfected using cotton wool soaked in 70% (v/v) ethanol and was allowed to run for at least five minutes before taking the sample into a sterile plastic container. The samples of sachet water were purchased from the retailers of the brands regularly patronized by the students. The water samples were transported to the laboratory in insulated containers with ice, stored in a refrigerator at a temperature at 4<sup>0</sup>C, and analyzed within twenty four hours of collection.

### **Physicochemical Analysis**

The water samples collected were analysed for pH, temperature, odour, colour, taste, turbidity, electrical conductivity, total dissolved solids, total alkalinity, total hardness, chloride, nitrate and selected trace metals.

### **Determination of pH**

The pH of the water samples was determined using the Wagtech pH meter. 10 ml of each of the samples was poured into a sterile beaker and the anode of the pH meter was dipped into it and readings were obtained when it was stable.

### **Measurement of Temperature**

This was conducted *in situ* at the site of sample collection using a mercury-in glass thermometer.

### **Determination of Colour, Odour and Taste**

A 20 mL volume of each water sample was poured into a clean beaker. The sample was then shaken vigorously to check for any frothing and allowed to settle. Colour was determined through visual examination and then odours were determined through the sensation of smell. Taste tests for consumer acceptability were performed on the water samples. Small volumes of each sample was tasted with the tongue and then immediately rinsed with distilled water (taste free) after each sample, and the results were recorded accordingly.

### **Determination of Turbidity**

A two part calibrated turbidity tube was used, with calibrations from 5-25 turbidity units. The joined tubes were held over a white paper, while slowly pouring the water sample into the tube until the black cross at the bottom was no longer visible. At this point the reading was taken from the side of the tube as the turbidity value of the water sample.

### **Determination of Electrical Conductivity**

A 20 mL volume of each water sample was poured into a clean beaker at room temperature. The Wagtech electrode was placed in the sample, and following stirring the electrical conductivity value was determined after the meter became stable.

### **Determination of Total Alkalinity**

10 ml of water sample was put in curvet; alkaphot tablet was added, crushed and mixed until all particles have dissolved. Optical density reading was taken at 570 nm wavelength on a Wagtech photometer.

### **Determination of Total Dissolved Solids**

The total dissolved solids were determined by submersing the Wagtech probe into 100 ml of each water sample following which the readings were recorded.

### **Determination of Chloride**

To fifty ml of the sample 5 drops of a phenolphthalein indicator solution was added and neutralized with 0.1N sulphuric acid to the colorless side of phenolphthalein. One ml of potassium chromate indicator solution was added before titration with standard silver nitrate solution to the pinkish-yellow endpoint. A reagent blank titration was carried out in parallel to the sample titration. Chloride concentration was calculated as follows (APHA, 2005): Chloride, mg/l =  $[(A-B) (N) (35.45)/V] \times 100$ , where A = Silver nitrate solution, in ml for sample titration; B = Silver nitrate solution, used for blank titration (in ml); N = Normality of the silver nitrate solution; and V = Sample volume in (ml).

### **Determination of Nitrate**

Test tube was filled with sample to 20 ml mark and one level spoonful

(~ 15 ml) of nitrate powder (containing zinc dust 60%, and barium sulphate 40%) and one nitrate tablet (ammonium chloride) were added and was shaken for a minute. The tube was allowed to stand for a minute and was inverted 3-4 times to aid flocculation and was allowed to stand for two minutes to ensure complete settlement. The clear solution was dispersed into tube to 10ml mark and one nitricol tablet (sulfanilic acid, acting as the aromatic amine), was added, crushed, and mixed to dissolve, then it was allowed to stand for 10 minutes for colour development and readings were taken on the photometer at 570 nm wavelength.

### **Determination of Total Hardness**

Total hardness was analyzed by titration of 50 ml water sample with standard EDTA

The EDTA was added in drops at pH 10 using Erichrome black T indicator until the colour changed into purple and the hardness was calculated by multiplying the average number of drops of EDTA used for the sample by the calibration factor of 20.

### **Determination of Iron**

A cuvette was filled with each water sample plus 0.25% ortho-phenanthroline solution (1:10 dilution), and optical density was taken at 510 nm wavelength using the Wagtech photometer. From a standard curve, the concentration of iron in the sample was determined.

### Determination of Zinc

Ten ml of water sample was dispensed in a test tube and one zinc tablet (5-(0-carboxyphenyl)-1-(2-hydroxy-5-sulphophenyl)-3-phenyl-formazan) was added, crushed, mixed to dissolve and was allowed to stand for five minutes and mixed to obtain a complete dissolution of the indicator. The optical density reading at 640 nm wavelength was taken using a Wagtech photometer.

### Bacteriological Analysis

Total heterotrophic bacteria in the water samples were obtained using the pour plate method. The enumeration and isolation of coliform bacteria was by the use of the membrane filtration technique (Eckner, 1998; Jagals *et al.*, 2000) and growth on MacConkey agar. The presence of *Escherichia coli* in the water samples was assessed by growth and colour reaction on Eosin Methylene Blue (EMB) agar, together with standard biochemical reactions as described (Barrow and Feltham, 1993).

### Results

The physicochemical properties of the water samples from boreholes, hand-dug wells and sachet water brands patronized by the students of Osun State University, main campus, residing in private hostels in Osogbo metropolis, examined in this study are as shown in Tables 1 and 2. All the water samples analyzed had similar and acceptable colour, odour and turbidity. The temperature of all

the samples were between 26.8 °C and 29.3 °C. The pH values of all the samples ranged between 6.1 and 8.4. The electrical conductivity, total dissolved solids and total hardness of all the water samples, ranged, respectively, between 97 and 305 µS/cm, 145 and 455 mg/l, and 84 and 446 mg/l. The total alkalinity was between 20 and 45 mg/l. The levels of zinc and chloride ranged, respectively, between 0.01 and 0.37 mg/l, and 5.0 and 32.0 mg/l while there were no detectable levels of arsenic ions in all the water samples analyzed. The tests for heavy metals such as lead, chromium or cadmium were not carried out in the present study. Two water samples in the borehole category (BH2 and BH3) had high levels of iron of 0.1 mg/l. Similarly, nitrate levels in two of the samples of water from hand-dug wells (HD1 and HD3) were determined to be greater than 50 mg/l.

The bacteriological properties of the water samples examined in this study are as shown in Table 3. Total viable bacterial counts were in the order of  $2.5 \times 10^1$  -  $7.0 \times 10^1$  colony forming units per ml of borehole and branded sachet water samples, and  $1.5 \times 10^3$  -  $2 \times 10^3$  in the water samples from hand-dug wells. Coliform counts per 100 ml of all water samples ranged from  $1.8 \times 10^1$  to  $1.7 \times 10^2$  while faecal coliform represented by *Escherichia coli* were 0.0 for all samples of borehole, branded sachet and one hand-dug

well (HD3) water, and 1.0 in two hand-dug wells (HD1 and HD2).

**Table 1: The pH, Conductivity, Total Alkalinity, Total Hardness and Total Dissolved Solids values of water samples from Osun State University, private student hostels, Osogbo**

<b>Water Sample</b>	<b>pH</b>	<b>Conductivity μS/cm</b>	<b>Total Alkalinity mg/l</b>	<b>Total Hardness mg/l</b>	<b>Total Dissolved Solids mg/l</b>
<b>BH1</b>	6.8	113	34	124	168
<b>BH2</b>	6.3	220	28	103	328
<b>BH3</b>	7.8	305	20	186	455
<b>HD1</b>	6.4	112	25	118	165
<b>HD2</b>	6.1	110	36	95	173
<b>HD3</b>	7.0	176	45	446	253
<b>SW1</b>	6.9	97	35	124	145
<b>SW2</b>	6.7	117	45	146	174
<b>SW3</b>	7.5	119	24	98	178
<b>SW4</b>	6.9	172	35	96	257
<b>SW5</b>	6.5	145	24	75	223
<b>SW6</b>	7.1	127	40	84	194
<b>SW7</b>	7.8	126	37	206	188
<b>SW8</b>	8.4	218	25	178	325
<b>SW9</b>	7.7	167	42	287	249
<b>WHO</b>	<b>6.5 – 9.2</b>	<b>-</b>	<b>120</b>	<b>500</b>	<b>1500</b>

<b>SON</b>	<b>6.5 - 8.5</b>	<b>1000</b>	<b>-</b>	<b>500</b>	<b>500</b>
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**Table 2: Metal ion (Arsenic, Iron and Zinc), Chloride and Nitrate values of water samples from Osun State University, private student hostels, Osogbo**

<b>Water Sample</b>	<b>Arsenic mg/l</b>	<b>Iron mg/l</b>	<b>Zinc mg/l</b>	<b>Chloride mg/l</b>	<b>Nitrate mg/l</b>
<b>BH1</b>	0.0	0.0	0.06	18	48
<b>BH2</b>	0.0	0.1	0.01	26	15
<b>BH3</b>	0.0	0.1	0.37	10	30
<b>HD1</b>	0.0	0.0	0.01	24	60
<b>HD2</b>	0.0	0.0	0.02	22	15
<b>HD3</b>	0.0	0.0	0.08	27	67
<b>SW1</b>	0.0	0.0	0.02	22	46
<b>SW2</b>	0.0	0.0	0.02	29	48
<b>SW3</b>	0.0	0.0	0.01	19	11
<b>SW4</b>	0.0	0.0	0.02	26	20
<b>SW5</b>	0.0	0.0	0.01	24	15
<b>SW6</b>	0.0	0.0	0.02	32	10
<b>SW7</b>	0.0	0.0	0.01	13	12
<b>SW8</b>	0.0	0.0	0.14	5	29
<b>SW9</b>	0.0	0.0	0.05	9	17



<b>WHO</b>	<b>0.01</b>	<b>0.3</b>	<b>4.0</b>	<b>250</b>	<b>50</b>
<b>SON</b>	<b>0.01</b>	<b>0.3</b>	<b>3.0</b>	<b>250</b>	<b>50</b>

**Table 3: Mean population of bacteria in water samples from Osun State University, private student hostels, Osogbo**

<b>Water Sample</b>	<b>Total bacterial count (cfu/ml)</b>	<b>Total coliform count (cfu/100ml)</b>	<b><i>Escherichia coli</i> (cfu/100ml)</b>
<b>BH1</b>	$2.5 \times 10^1$	$4.5 \times 10^1$	0.0
<b>BH2</b>	$6.0 \times 10^1$	$1.1 \times 10^2$	0.0
<b>BH3</b>	$7.0 \times 10^1$	$1.3 \times 10^2$	0.0
<b>HD1</b>	$2.3 \times 10^3$	$1.7 \times 10^2$	1.0
<b>HD2</b>	$1.9 \times 10^3$	$1.6 \times 10^2$	1.0
<b>HD3</b>	$1.5 \times 10^3$	$1.4 \times 10^2$	0.0
<b>SW1</b>	$2.5 \times 10^1$	$2.4 \times 10^1$	0.0
<b>SW2</b>	$2.0 \times 10^1$	$3.2 \times 10^1$	0.0
<b>SW3</b>	$2.7 \times 10^1$	$1.8 \times 10^1$	0.0
<b>SW4</b>	$3.5 \times 10^1$	$5.4 \times 10^1$	0.0
<b>SW5</b>	$5.3 \times 10^1$	$6.5 \times 10^1$	0.0
<b>SW6</b>	$7.0 \times 10^1$	$6.1 \times 10^1$	0.0
<b>SW7</b>	$4.8 \times 10^1$	$7.3 \times 10^1$	0.0
<b>SW8</b>	$6.5 \times 10^1$	$8.0 \times 10^1$	0.0

<b>SW9</b>	4.2 x 10 <sup>1</sup>	6.3 x 10 <sup>1</sup>	0.0
<b>WHO</b>			0.0
<b>SON</b>		1.0 x 10 <sup>3</sup>	0.0

## Discussion

This study determined the quality of water from borehole, hand-dug wells and sachet water brands patronized by the students of Osun State University, main campus, residing in private hostels in Osogbo metropolis. Virtually all of the samples of sachet water examined in the study exhibited values of physical / organoleptic parameters, inorganic constituents, and mean coliform and *E. coli* counts per 100 ml below the WHO/SON (WHO, 2007; SON, 2007) maximum permissible levels; and are therefore, considered safe for drinking. The results of the physicochemical and bacteriological assessment of the collected sachet water samples confirmed with the work done earlier in Osogbo (Olowe *et al.*, 2005), Ilorin (Adewoye *et al.*, 2011), Ota (Chinedu *et al.*, 2011), and Accra, Ghana (Addo *et al.*, 2009) and this could be attributed to the total adherence to strict quality assurance procedures in the laboratory and production premises of the companies concerned. It is noteworthy, however, that some assessment of sachet water brands vended in Ondo (Onifade and Ilori, 2008), Kebbi (Kalpana *et al.*, 2011) Ogbomoso (Oladipo *et al.*, 2009) and

Ibadan (Adekunle *et al.*, 2004; Ajayi *et al.*, 2008) have physicochemical values of concern and/or evidence of faecal coliforms. These contradictory reports highlight the need for ongoing implementation of the several legislations put in place by National Agency for Drug Administration and Control (NAFDAC) in Nigeria, and continuous monitoring to increase reduction in the sales of contaminated brands of sachet water; and this may be the reflection of what we have observed in this work.

The presence of iron in groundwater supplies is a common problem (Merrill *et al.*, 2009) The slightly elevated levels of iron in water samples from the borehole category, BH2 and BH3 may have arisen naturally in the aquifer, and its levels could also be increased by dissolution of ferrous borehole or by the action of iron related bacteria. Interestingly, no health-based value for iron in drinking water has been proposed in the WHO guidelines (WHO, 2003). Although, iron stains laundry at levels above 0.3 mg/l, there is usually no noticeable taste in iron concentration below 0.3 mg/l (WHO, 2003). The acceptability of water samples from the boreholes

examined in this study was not questionable for drinking purposes, given the mean coliform counts per 100 ml below the WHO/SON maximum permissible levels and lack of faecal *E. coli* in the samples. This work confirms the work done by Yusuf *et al.*, (2012) which reported that all the borehole water sampled in Offa and Erin-Ile in Kwara State and in Ilesa and Osogbo in Osun State, Nigeria were free of *E. coli*. In a study conducted by Akpoveta *et al.*, 2011, physicochemical parameters determined for borehole water used in the vicinities of Benin, Edo State and Agbor, Delta State of Nigeria were found to fall within the WHO/SON maximum permitted limit for potable water except for calcium and manganese.

The slightly elevated nitrate levels in samples from hand-dug wells, HD1 and HD3, and evidence of faecal coliform in samples from hand-dug wells HD1 and HD2 suggest that these water sources may have been contaminated as a consequence of agricultural activity (including excess application of inorganic nitrogenous fertilizers and manures), from wastewater disposal and from oxidation of nitrogenous waste products in human and animal excreta, including septic tanks that were located in the close vicinity of the wells. According to WHO (2007), nitrate may have a role in protecting the gastrointestinal tract against a variety of gastrointestinal

pathogens since nitrous oxide and acidified nitrite have antibacterial properties, however, nitrate in drinking water is an important risk factor for bottle-fed infants (under three months) given increases in methaemaglobinaemia and possible cyanosis (Greer and Shannon, 2005). Older children are relatively insensitive to the effects of nitrate (Craun, Greathouse & Gunderson, 1981). A number of studies (Gatseva & Argirova, 2008; Radikova *et al.*, 2008; Ward *et al.*, 2010) have reported a correlation between various measures of nitrate intake and effects on thyroid function, but all suffer from methodological and data problems that preclude definitive conclusions being drawn about the effect of nitrates in adults. Practical considerations to control nitrate concentration in well water include appropriate management of agricultural practices, well thought-out siting of pit latrines and septic tanks, sewer leakage control, as well as management of fertilizer and manure applications (WHO, 2007).

*E. coli* may have entered the private wells through contamination with faecal matter from infested or infected humans or animals, through sewage overflows, polluted storm water runoff, and agricultural runoff. In addition, the wells may have been more vulnerable to such contamination after flooding, since most hand-dug wells are usually shallow (Ayantobo *et al.*, 2013). The evidence for *E. coli* in two of three

hand-dug wells examined in this study raises concern given the rise of infection rate of *E. coli* O157:H7, a potentially life-threatening pathogen, in Southwest Nigeria where 4.3% out of one thousand and eight hundred diarrhoeic patients sampled in a recent study were positive for the organism, the highest occurrence (4.5%) was found among those using well water, and 5.8% among those people using dunghill as toilet (Ademokoya *et al.*, 2013). Boil water orders, water avoidance advisories, proper excreta disposal and improvement in general hygiene would enhance the quality of *E. coli* infested well water sources and reduce possible infection by the indicator bacteria.

We conclude, on the bases of the physicochemical and bacteriological properties of the water samples examined in this study that water

from boreholes and sachet water brands patronized by the students of Osun State University, main campus, residing in private hostels in Osogbo metropolis are safe for human consumption. While the water samples from hand-dug wells may be useful for other purposes such as cleaning, bathing, and toilet flushing, they are not safe for consumption by infants under three months old. Because of the presence of *E. coli* in some well water samples it is recommended that water from these sources should be boiled before consumption to ensure public health and safety.

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